

HIV Through the Looking Glass: Insights Derived From Hepatitis B

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Background: Although higher levels of hepatitis B virus (HBV) replication in HIV-HBV co-infection may relate to liver disease progression, this has not been completely elucidated. We used expression of hepatitis B core antigen (HBcAg) in liver biopsies from HIV-HBV co-infected and HBV mono-infected patients as a marker for HBV replication, and related these findings to clinical and histological parameters.

Methods: Data from 244 HBV patients were compared with 34 HIV-HBV patients. Liver biopsies were scored for inflammation, fibrosis, HBcAg, and hepatitis B surface antigen. Univariate and multivariate analyses were performed.

Results: HBcAg, but not hepatitis B surface antigen, staining was stronger in HIV co-infected than in HBV mono-infected. Co-infected and HBV mono-infected had similar alanine aminotransferase, inflammatory and fibrosis scores, and hepatitis B e antigen status. HBcAg staining correlated with HIV after correcting for HBV DNA and hepatitis B e antigen. CD4 counts and HIV RNA level did not correlate with intensity of HBcAg staining. HBV DNA levels were higher in HIV co-infected and correlated with HBcAg staining.

Conclusions: By looking at HBcAg as a reflection of HBV replication in HIV-HBV co-infected with controlled HIV, our

findings suggest that these patients may have subtle immune function defects, which could lead to adverse liver disease outcomes.

Key Words: co-infection, HBV, HIV, immune

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INTRODUCTION

Persons with HIV infection are not infrequently co-infected with hepatitis B virus (HBV), largely because these 2 viruses share similar modes of transmission. The frequency of HBV co-infection in HIV-infected persons in the United States ranges from 5% to 10% but is higher in the areas of the world where the background rates of HBV infection are higher and ranges from 20% to 30% in Asia and sub-Saharan Africa.¹

Co-infection with HBV is important in the natural history and outcome of chronic HIV infection. In recent years, liver disease has become a primary cause of disease progression, morbidity, and mortality in persons with AIDS. Thus, chronic HIV infection seems to worsen the natural history and outcome of hepatitis B; and in return, HBV exacerbates the course of chronic HIV infection. In addition, the mortality rate of chronic HIV infection is increased in those with HBV co-infection.^{2–4}

The interactions between HBV and HIV infection that lead to worse clinical outcomes of both viruses are not completely understood. A major element seems to be suppression of the immune response because of HIV infection causing an increase in HBV replication, which in turn causes worsening of the accompanying liver disease. In view of this, one might expect that improvement in therapies of HIV infection would improve the outcome of hepatitis B in co-infected individuals. However, with introduction of highly active antiretroviral therapy, the rates of death because of liver disease in HIV-infected cohorts seemed to increase, and it was only with satisfactory therapies of the concomitant hepatitis B using agents, which were active against both HIV and HBV, that improved morbidity and mortality could be demonstrated.^{5–7}

HIV-HBV co-infected patients with controlled HIV disease (relatively low viral load and relatively high CD4 count) were observed to have liver biopsies with more widespread staining for hepatitis B core antigen (HBcAg) staining, a marker of active HBV replication, compared with

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those of HBV mono-infected patients. Patients with well-controlled HIV would be expected to have HBV levels within the normal range. However, this higher level of HBcAg staining suggested that HBV disease was not controlled. We hypothesized that increased staining seen in HIV-HBV co-infection was related to facilitated HBV replication in patients with a compromised immune system because of HIV. The aim was to identify factors that correlated with HBcAg and hepatitis B surface antigen (HBsAg) staining in hepatocytes of patients with HIV-HBV co-infection, focusing on the serum and tissue markers of HBV and HIV infection.

METHODS

Patients

Liver biopsies performed between 1980 and 2002 on adult patients with chronic hepatitis B by the members of the Liver Diseases Branch of the Clinical Center of the NIH were selected for review. Patients with concomitant liver diseases were excluded as were those who were receiving HBV antiviral therapy at the time of liver biopsy. Clinical and laboratory data were extracted from patient charts. In patients without available results, stored serum samples taken at or around the time of liver biopsy (within 2 months) were retrieved and tested for HBV DNA. Patients with insufficient clinical data were excluded from analysis.

HBV DNA levels were measured in stored serum samples by quantitative polymerase chain reaction (Roche COBAS TaqMan HBV Analyte Specific Reagent). HIV RNA levels were measured by Roche COBAS Amplicor HIV-1 Monitor Test. HIV antibodies were measured by Ortho VITROS Anti-HIV 1 + 2 assay. Only the first liver biopsy was included in patients with more than 1 assessment.

All patients studied were participants in clinical research studies being conducted at the National Institute of Allergy and Infectious Diseases and National Institute of Diabetes and Digestive and Kidney Diseases of the National Institutes of Health and gave written informed consent for studies of the HIV and/or HBV infection.

Liver Histology

Liver histology was evaluated by a hepatic pathologist without knowledge of the clinical features of the patients. Inflammatory activity was graded using a modification of the histology activity index, and fibrosis was staged using the Ishak fibrosis score. All liver biopsies were also read for the presence of HBcAg and HBsAg by immunoperoxidase staining and scored without knowledge of HIV status. The degree of HBV antigen staining was scored on a scale of 0–4+ based on the proportion of hepatocytes with positive staining in which 0 = none, 1+ = <10%, 2+ = 10%–50%, 3+ = 50%–90%, and 4+ = >90% stained. The pattern of distribution of HBcAg was also recorded as either nuclear only, nuclear predominant, mixed nuclear and cytoplasmic, and cytoplasmic only.

Statistical Analysis

Univariate analyses were performed with *t* tests, Mann–Whitney, or χ^2 tests, as appropriate, using a $P < 0.05$ as significant. Multivariate analysis included variables with a $P < 0.10$ in univariate analysis. Multicollinearity among variables included in the model was defined as an $R^2 \geq 0.75$ between any 2 variables.

RESULTS

Between 1980 and 2002, 823 liver biopsies were performed at the National Institutes of Health in patients with chronic hepatitis B. Only 369 biopsies, however, were the initial biopsy and were from adult patients who were not already on HBV antiviral therapy. After initial analysis of these subjects, 83 were excluded because of insufficient clinical information and 8 because of a concurrent liver disease including hepatitis C ($n = 6$), chronic alcoholism ($n = 1$), and primary sclerosing cholangitis ($n = 1$). Of the remaining 278 patients, 34 (12%) were co-infected with HIV. A comparison of the demographic, clinical, laboratory, and histologic characteristics of patients with HBV mono-infection versus HIV-HBV co-infection are shown in Table 1. Patients with HIV-HBV co-infection were statistically significantly more likely to be male (100% vs 80%) and white (95% vs 64%) than those with HBV mono-infection. The HIV-infected cohort was also slightly younger (39 vs 42 years), although the known duration of HBV infection was longer (9 vs 2.5 years), than those with HBV alone. Nevertheless, serum alanine aminotransferase (ALT) levels, platelet counts, and liver histology scores were similar between the 2 groups ($P > 0.05$ with all 3 parameters).

In contrast to the clinical and biochemical features, the viral factors were different between the 2 groups. HBV DNA levels were higher in co-infected patients (median = 4.2 billion vs 526 million copies per milliliter), and they were more likely to have hepatitis B e antigen (HBeAg) in serum (96% vs 80%). Additionally, staining patterns differed between the 2 groups (see **Figure S1, Supplemental Digital Content**, <http://links.lww.com/QAI/A589>). HBcAg staining was more frequent (22.7 vs 5.6%) and more intense (mean staining scores = 1.7 vs 2.5) in the HIV-HBV co-infected than the HBV mono-infected group (see **Figure S2A, Supplemental Digital Content**, <http://links.lww.com/QAI/A589>). Even with adjustments for gender, age, race, total inflammatory score, and Ishak fibrosis score, the co-infected group had a higher odds ratio for both HBc staining (odds ratio = 3.71, $P = 0.01$) and HBV DNA (odds ratio = 1.75, $P < 0.01$) (Table 2). The pattern of staining was most often “mixed nuclear and cytoplasmic” in the HIV-HBV co-infected and more frequently “nuclear only” in the HBV mono-infected subjects. In contrast, HBsAg staining was similar in the 2 groups, with mean staining score 1.1 vs 1.4 (see **Figure S2B, Supplemental Digital Content**, <http://links.lww.com/QAI/A589>).

Associations were sought between HIV status and clinical features, as well as biochemical and HBV virologic factors. The majority of HIV-infected subjects had reasonable immunologic function. CD4 counts were >200 cells per

TABLE 1. The Demographic, Clinical, Laboratory, and Histologic Characteristics of HBV Mono-infected and HIV-HBV Co-infected Patients

Variable	HBV		HIV-HBV		P
	n	%	n	%	
Total	195		22		
Sex					
Male	155	79.5	22	100.0	0.017
Female	40	20.5	0	0.0	
Race					
White	125	64.1	21	95.5	0.015
Black	17	8.7	1	0.5	
Asian	43	22.1	0		
Other/not available	10	5.1	0		
Age (mean ± SD), yrs	187	41.6 ± 12.9	22	38.8 ± 5.8	0.077
BMI (mean ± SD), kg/m ²	144	25.5 ± 4.2	16	25.2 ± 3.3	0.820
Time from HBV diagnosis (median), yrs	164	2.5	22	9.3	<0.001
ALT (median), IU/dL	128	97.5	21	143	0.226
Platelet count (median), platelets/mm ³	115	183	22	159	0.179
HBV DNA (median), copies/mL	526,333,000		4,155,000,000		0.019
HBeAg (% positive)	80		96		0.051
HIV viral load (%)					
<50, copies/mL	—		63		—
50–10,000, copies/mL	—		25		—
>10,000, copies/mL	—		12		—
CD4 count (mean ± SD), cells/mm ³	—		468 ± 270		—
CD4 count (% >200 cells/mm ³)	—		81.3		—
Periportal (mean ± SD, scale 0–10)	195	3.8 ± 2.1	22	3.7 ± 1.9	0.860
Total inflammatory (mean ± SD, scale 0–18)	195	9.0 ± 3.2	22	8.7 ± 3.7	0.631
Ishak fibrosis score (mean ± SD, scale 0–6)	192	2.9 ± 1.8	22	3.1 ± 1.4	0.643
HBsAg (mean ± SD, scale 0–4)	1.1 ± 0.8		1.4 ± 1.1		0.22
HbCg (mean ± SD, scale 0–4)	1.65 ± 1.1		2.5 ± 1.2		0.0044
HbCg pattern					
1 = Nuclear only	36%		11%		0.0231
2 = Nuclear predominant	28%		26%		
3 = Mixed, nuclear and cytoplasmic	26%		58%		
4 = Cytoplasmic only	10%		5%		

The mean was compared with *t* test, the medians were compared with Wilcoxon 2-sample test, and the proportions were compared with Fisher exact test.

cubic millimeter in 81%, and HIV viral loads were <10,000 copies per milliliter in 84% of patients. Nonetheless, these HIV-positive patients had higher levels of HBV DNA and more intense HbCg in liver tissue than the mono-infected patients despite similar degrees of disease activity as shown by serum ALT levels and histology activity index scores, as well as similar degrees of disease stage as shown by platelet counts and fibrosis scores (see **Figure S3A, Supplemental Digital Content**, <http://links.lww.com/QAI/A589>). Indeed, peripheral blood CD4 counts and HIV RNA levels did not correlate with HBV DNA levels nor with the intensity of HbCg staining in the 18 patients with available data (all *P* > 0.20) (see **Figure S3B and S3C, Supplemental Digital Content**, <http://links.lww.com/QAI/A589>).

HBV DNA levels correlated strongly with degree of HbCg staining and were higher in those with HBeAg in serum. However, even after controlling for HBV DNA levels

and HBeAg status, HbCg staining intensity was still greater in HIV co-infected patients than in HBV mono-infected patients (*R*² = 0.36; *P* < 0.0001). Examples of typical HbCg staining pattern in an HBV mono-infected patient compared with such typical staining in an HIV-HBV co-infected patient are shown in **Figure S3** (see **Supplemental Digital Content**, <http://links.lww.com/QAI/A589>).

DISCUSSION

High serum HBV DNA levels, positive HBeAg, positive HbCg, and increased serum ALT levels are signs of active HBV replication. In HBV mono-infection, active HBV replication is linked with increased severity of liver disease and liver-related mortality.^{8–11}

In our study cohort, HBV-HIV co-infected patients had CD4 counts mostly above 200, low HIV RNA levels, and

TABLE 2. Odds Ratios of HBcAg in Relationship to HIV Status and HBV DNA

Variable	Odds Ratio	95% CI	P
Group			
HBV	Ref.		
HIV-HBV	3.71	1.34 to 10.27	0.012
DNA			
Log 10	1.75	1.44 to 2.12	<0.001

Outcome: HBcAg (ordered value, according to the levels of staining).
Number of subjects included in analysis: 134.
Adjusted for gender, age, race, total inflammatory score, and Ishak fibrosis score.

similar ALT, platelet counts, inflammation and fibrosis scores when compared with HBV mono-infection. Co-infected patients had higher serum HBV DNA and increased HBcAg hepatocyte staining despite low HIV RNA levels and independent of CD4 counts, serum HBeAg status, ALT level, or hepatic inflammation. HBcAg localization has been previously linked to more severe disease.^{12–14} Showing increased HBcAg supports the notion that HBV was actively replicating at a higher rate in HIV-HBV co-infected than mono-infected despite no indications of immunosuppression or advanced disease. This implies that subtle defects by the host immune system in even well-controlled HIV lead to enhanced HBV replication.

Our work adds to other previously described work that suggested functional lymphocyte abnormalities and lower T4/T8 ratios in patients with HIV infection compared with those without the infection may help explain the subtle deficits.^{15,16} A recent, large retrospective study from Taiwan looked at the correlation between HIV-positive patients and hepatitis B and hepatitis C serum markers.¹⁷ Our work expands their chart review with liver biopsy observations. Although we did not find an association between CD4 counts and anti-HBcAg staining, our findings, in particular that HBcAg staining is correlated with HIV co-infection, are consistent with their conclusion that compromised immunity affects HBV markers.

Although increased HBV DNA and HBcAg expression does not directly measure disease severity, it does offer insights into how the disease state in HIV-HBV co-infection is altered compared with HBV alone. Despite what seems to be controlled HIV disease, HBV liver disease remains uncontrolled. Although studies have reported that liver disease progression is accelerated in HIV-HBV co-infection, the reason for this is still not well understood. Studies have shown that HIV-infected individuals possess features characteristic of an aging immune system or “immune senescence” that is driven by chronic immune activation.¹⁸ In another study looking at CD4 T-cell decline in pediatric HIV-1 infection, microbial translocation was associated with persistent monocyte/macrophage activation unrelated to viral replication or T-cell activation.¹⁹ Similarly, microbial translocation has been linked to hepatitis C virus liver disease progression in patients co-infected with HIV.²⁰ It is important to explore these and other possible mechanisms because it affects clinical outcomes.

We acknowledge several limitations to our retrospective study. After exclusion criterion, the sample size of co-infected patients was relatively small. Polymerase chain reaction detection of viral nucleic acid levels at a single time point may encompass transient variations. However, we do not believe these variables significantly bias our results.

In summary, using liver biopsies from HBV or HIV-HBV patients, we find significantly increased HBcAg but not HBsAg staining in co-infected patients, despite similar fibrosis levels. We also find that HBV DNA levels were higher in co-infected patients, although CD4 counts were independent of HBV DNA levels.

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REFERENCES

- Koziel MJ, Peters MG. Viral hepatitis in HIV infection. *N Engl J Med*. 2007;356:1445–1454.
- Hadler SC, Judson FN, O'Malley PM, et al. Outcome of hepatitis B virus infection in homosexual men and its relation to prior human immunodeficiency virus infection. *J Infect Dis*. 1991;163:454–459.
- Thio CL, Seaberg EC, Skolasky R Jr, et al. HIV-1, hepatitis B virus, and risk of liver-related mortality in the Multicenter Cohort Study (MACS). *Lancet*. 2002;360:1921–1926.
- Spradling PR, Richardson JT, Buchacz K, et al. Prevalence of chronic hepatitis B virus infection among patients in the HIV Outpatient Study, 1996–2007 (dagger). *J Viral Hepat*. 2010;17:879–886.
- Thio CL. Hepatitis B in the human immunodeficiency virus-infected patient: epidemiology, natural history, and treatment. *Semin Liver Dis*. 2003;23:125–136.
- Konopnicki D, Mocroft A, de Wit S, et al. Hepatitis B and HIV: prevalence, AIDS progression, response to highly active antiretroviral therapy and increased mortality in the EuroSIDA cohort. *AIDS*. 2005;19:593–601.
- Hoffmann CJ, Seaberg EC, Young S, et al. Hepatitis B and long-term HIV outcomes in coinfecting HAART recipients. *AIDS*. 2009;23:1881–1889.
- Benhamou Y. Hepatitis B in the HIV-coinfected patient. *J Acquir Immune Defic Syndr*. 2007;45(suppl 2):S57–S65; discussion S66–57.
- Iloeje UH, Yang HI, Jen CL, et al. Risk and predictors of mortality associated with chronic hepatitis B infection. *Clin Gastroenterol Hepatol*. 2007;5:921–931.
- Ramalho F, Brunetto MR, Rocca G, et al. Serum markers of hepatitis B virus replication, liver histology and intrahepatic expression of hepatitis B core antigen. *J Hepatol*. 1988;7:14–20.
- McDonald JA, Harris S, Waters JA, et al. Effect of human immunodeficiency virus (HIV) infection on chronic hepatitis B hepatic viral antigen display. *J Hepatol*. 1987;4:337–342.
- Hsu HC, Su IJ, Lai MY, et al. Biologic and prognostic significance of hepatocyte hepatitis B core antigen expressions in the natural course of chronic hepatitis B virus infection. *J Hepatol*. 1987;5:45–50.
- Naoumov NV, Portmann BC, Tedder RS, et al. Detection of hepatitis B virus antigens in liver tissue. A relation to viral replication and histology in chronic hepatitis B infection. *Gastroenterology*. 1990;99:1248–1253.
- Chu CM, Liaw YF. Intrahepatic distribution of hepatitis B surface and core antigens in chronic hepatitis B virus infection. Hepatocyte with cytoplasmic/membranous hepatitis B core antigen as a possible target for immune hepatocytolysis. *Gastroenterology*. 1987;92:220–225.

15. Krohn K, Ranki A, Anttonen J, et al. Immune functions in homosexual men with antibodies to HTLV-III in Finland. *Clin Exp Immunol*. 1985;60:17–24.
16. Nicholson JK, McDougal JS, Jaffe HW, et al. Exposure to human T-lymphotropic virus type III/lymphadenopathy-associated virus and immunologic abnormalities in asymptomatic homosexual men. *Ann Intern Med*. 1985;103:37–42.
17. Sun HY, Lee HC, Liu CE, et al. Factors associated with isolated anti-hepatitis B core antibody in HIV-positive patients: impact of compromised immunity. *J Viral Hepat*. 2010;17:578–587.
18. Nixon DE, Landay AL. Biomarkers of immune dysfunction in HIV. *Curr Opin HIV AIDS*. 2010;5:498–503.
19. Wallet MA, Rodriguez CA, Yin L, et al. Microbial translocation induces persistent macrophage activation unrelated to HIV-1 levels or T-cell activation following therapy. *AIDS*. 2010;24:1281–1290.
20. Balagopal A, Philp FH, Astemborski J, et al. Human immunodeficiency virus-related microbial translocation and progression of hepatitis C. *Gastroenterology*. 2008;135:226–233.